

Store at  
4°C

# Protein L (PE Conjugate)



Cell Signaling  
TECHNOLOGY®

#58036

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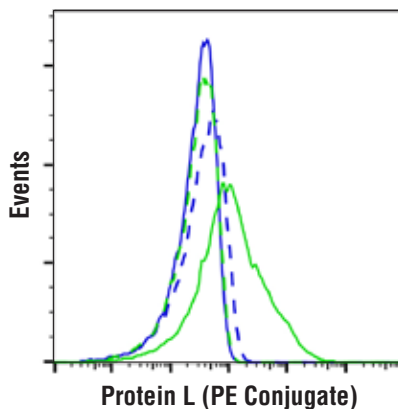
## For Research Use Only. Not For Use In Diagnostic Procedures.

**Description:** This Cell Signaling Technology product is conjugated to phycoerythrin (PE) and tested in-house for direct flow cytometric analysis in human cells.

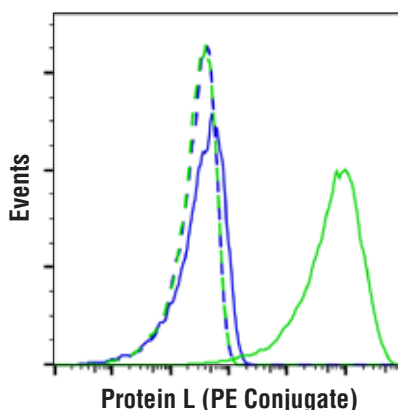
**Background:** Protein L was first isolated from the surface of bacterial species *Peptostreptococcus magnus* (1). Unlike Protein A and Protein G that differentially bind to the Fc region of immunoglobulin isotype heavy chains, Protein L contains Ig binding domains that interact with the  $\kappa$  light chain variable domain without disrupting paratope function (2-4). This attribute not only facilitates binding to a broader range of immunoglobulin classes and subclasses than either Protein A or Protein G but also allows for binding to Fab fragments and single chain variable fragments (scFvs) that contain  $\kappa$  light chain sequences. Protein L does, however, display specificity with respect to  $\kappa$  light chain subtypes. In human, Protein L binds to V $\kappa$ I, V $\kappa$ II, and V $\kappa$ IV subtypes whereas binding is restricted to V $\kappa$ I in mice (4).

**Specificity/Sensitivity:** Conjugation to PE makes this product ideally suited for direct flow cytometric analysis of cells expressing  $\kappa$  light chain-containing immunoglobulins as well as immunoglobulin fragments that contain  $\kappa$  light chains, such as scFvs.

**Source/Purification:** Protein L (PE Conjugate) is recombinantly derived and genetically engineered such as to remove the cell wall-, cell membrane-, and albumin-binding regions of Protein L to enhance binding to immunoglobulins. The purified Protein L was conjugated under optimal conditions, with unreacted dye removed from the preparation.



Flow cytometric analysis of live Jurkat cells (blue) or live Jurkat cells engineered to stably express an scFv-based Anti-CD19 CAR (green), using Protein L (PE Conjugate) (solid lines) or unstained cells (dashed lines). Cell line was provided by the Lohmueller Lab, University of Pittsburgh.



Flow cytometric analysis of live Jurkat cells (blue) or live Jurkat cells engineered to stably express an scFv-based Anti-CD20 CAR (green), using Protein L (PE Conjugate) (solid lines) or unstained cells (dashed lines). Cell line was provided by the Lohmueller Lab, University of Pittsburgh.

**Storage:** Supplied in PBS (pH 7.2), less than 0.1% sodium azide and 2 mg/ml BSA. Store at 4°C. Do not aliquot. Protect from light. Do not freeze.

**Directions for Use:** Recommended Dilution: 1:50

### Background References:

- (1) Liebermann, B. et al. (1990) *Dtsch Z Mund Kiefer Gesichtschir* 14, 418-23.
- (2) Nilson, B.H. et al. (1992) *J Biol Chem* 267, 2234-9.
- (3) Akerström, B. and Björck, L. (1989) *J Biol Chem* 264, 19740-6.
- (4) Nilson, B.H. et al. (1993) *J Immunol Methods* 164, 33-40.

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**Applications:** W—Western IP—Immunoprecipitation IHC—Immunohistochemistry CHIP—Chromatin Immunoprecipitation IF—Immunofluorescence F—Flow cytometry E-P—ELISA-Peptide **Species Cross-Reactivity:** H—human M—mouse R—rat Hm—hamster Mk—monkey Mi—mink C—chicken Dm—D. melanogaster X—Xenopus Z—zebrafish B—bovine Dg—dog Pg—pig Sc—S. cerevisiae Ce—C. elegans Hr—Horse All—all species expected Species enclosed in parentheses are predicted to react based on 100% homology.

## Flow Cytometry, Live Cell Protocol for Directly Conjugated Protein L

### A. Solutions and Reagents

**NOTE:** Prepare solutions with reverse osmosis deionized (RODI) or equivalent grade water.

- 1X Phosphate Buffered Saline (PBS):** To prepare 1 L 1X PBS: add 100 ml 10X PBS (#12528) to 900 ml water, mix.
- Dilution Buffer:** Purchase ready-to-use Flow Cytometry Antibody Dilution Buffer (#13616) that is compatible with Protein L, or prepare a 0.5% BSA PBS buffer by dissolving 0.5 g Bovine Serum Albumin (BSA) (#9998) in 100 ml 1X PBS. Store at 4°C.

**NOTE:** When including fluorescent cellular dyes in your experiment (including viability dyes, DNA dyes, etc.), please refer to the dye product page for the recommended protocol. Visit [www.cellsignal.com](http://www.cellsignal.com) for a full listing of cellular dyes validated for use in flow cytometry.

### B. Immunostaining

**NOTE:** Count cells using a hemocytometer or alternative method.

**NOTE:** If using whole blood, lyse red blood cells and wash by centrifugation prior to Immunostaining.

**NOTE:** Optimal centrifugation conditions will vary depending upon cell type and reagent volume. Generally, 150-300g for 1-5 minutes will be sufficient to pellet the cells.

- Aliquot desired number of cells into tubes or wells. (Generally,  $5 \times 10^5$  to  $1 \times 10^6$  cells per assay.)
- Pellet cells by centrifugation and remove supernatant.
- Resuspend cells in 100  $\mu$ l of diluted Protein L conjugate, prepared in Dilution Buffer at a recommended dilution or as determined via titration.
- Incubate for 30 min to 1 hr on ice. Protect from light.
- Wash by centrifugation in Dilution Buffer. Discard supernatant. Repeat. If using a fluorescent conjugate of Protein L, skip to step 9.
- For labeling biotinylated Protein L, resuspend cells in 100  $\mu$ l of diluted fluorochrome-conjugated streptavidin or anti-biotin antibody (prepared in Dilution Buffer at the recommended dilution).
- Incubate for 30 min on ice. Protect from light.
- Wash by centrifugation in Dilution Buffer. Discard supernatant. Repeat.
- Resuspend cells in 200-500  $\mu$ l of Dilution Buffer and analyze on a flow cytometer.